

0960-894X(95)00585-4

SYNTHESIS, ENZYMATIC PHOSPHORYLATION AND ANTIVIRAL ACTIVITY OF ACYCLIC DIENYL PHOSPHONATE DERIVATIVES OF GUANINE

Jean-François Navé*+, Patrick J. Casara+, Debra L. Taylor#, A. Stanley Tyms#,
Michael Kenny† and Serge Halazy+¤

⁺Marion Merrell Dow Research Institute, 16 rue d'Ankara, 67080 Strasbourg, France

[†] Marion Merrell Dow Inc., 10236 Marion Park Drive, Kansas City, MO, USA

[#]MRC, Collaborative Centre, 1-3 Burtonhole Lane, Mill Hill, London, UK

^{**}Present address: Centre de Recherche Pierre Fabre, 17 Avenue Jean Moulin, 81106 Castres, France

Abstract: The synthesis, phosphorylation by guanylate kinase, anti HIV-1 and anti-herpesvirus activity of two acyclic dienyl phosphonate derivatives of guanine are described. (E)-9-(5-phosphono-3-methylene-4-pentenyl)guanine (4) was identified as an excellent substrate of guanylate kinase and a significant inhibitor of HIV-1 replication.

In recent years, numerous acyclonucleotide analogues have been prepared as potential antiviral agents ¹⁻⁶. These compounds bear a chemically stable phosphonate-containing moiety in place of the labile phosphate monoester group and are structural mimics of nucleoside monophosphates. It has been shown that various acyclonucleotide analogues exert their antiviral effect following sequential activation by cellular kinases to their diphosphate derivatives ^{7,8} (nucleoside triphosphate analogs) which act as inhibitors of viral DNA polymerases ^{1,7,8}. As mimics of the phosphate monoester group, the phosphonomethyl and especially the phosphonomethoxy groups have been used. This has led to the discovery of promising antiviral agents such as 9-[2-(phosphonomethoxy)ethyl]adenine (PMEA, Adefovir)⁹ and (S)-1-(3-hydroxy-2-phosphonomethoxypropyl)-cytosine ((S)-HPMPC, Cidofovir)⁹. Very recently, acyclonucleotide analogues bearing an α , β -unsaturated phosphonate moiety as mimic of the phosphate group were found to exhibit significant antiviral activity ^{10,11}. Interestingly, we noticed that unlike (E)-9-[(2-hydroxymethyl-4-phosphonobut-3-enyl)oxy]guanine (1) (Scheme 1), the carba analogue 2 had low cellular toxicity while retaining anti-HIV activity. Moreover, compound 2 was found to be a good substrate of guanylate kinase ¹¹, an enzyme held responsible for the intracellular phosphorylation of the monophosphates of acyclovir and ganciclovir ^{12,13} and more generally of various acyclic phosphonate derivatives of guanine ^{1,4,11}. These results suggested that the dienyl phosphonate 4 (scheme 1)

Scheme 1

could also be an efficient substrate of guanylate kinase. If so, and provided that the product of this first step is further phosphorylated by known cellular kinases^{1,14} of broad specificities, the diphosphate of **4** could be formed intracellularly and perhaps behave as a good inhibitor of viral DNA polymerases. We report here the synthesis of **4** and of its shorter chain analogue **3**, their substrate properties for guanylate kinase as well as their antiviral activity profile. While this work was in progress, Harnden and collaborators¹⁰ reported the synthesis of a series of phosphonoalkenyl and phosphonoalkenyloxy derivatives of purines. During that work, the dienyl phosphonate **3** was obtained incidentally and found to be inactive both against HIV-1 and herpes viruses¹⁰.

Syntheses of (E)-9-(4-phosphono-2-methylene-3-butenyl)guanine **3** and (E)-9-(5-phosphono-3-methylene-4-pentenyl)guanine **4** were performed as outlined in scheme 2, starting from 3-t-butyldimethylsilyloxy-2-methylene-propanal **5** and 4-t-butyldimethylsilyloxy-3-methylene-butanal **6**, respectively. Condensation of these aldehydes with the lithium salt of tetraethyl methylene-bis-phosphonate¹⁵ gave the corresponding dienylphosphonates **7** and **8**. The t-butyldimethylsilyl groups were removed to give the corresponding hydroxy derivatives which were converted to the sulfonates **9** and **10**. The purine moiety was then introduced by alkylation of **9** or **10** with the sodium salt of 6-chloro-2-aminopurine in acetonitrile to give the corresponding protected nucleotides **11** and **12**. These products were obtained with a high regioselectivity (N9/N7 > 90% as determined by 1 H NMR spectroscopy). The desired acyclonucleotides 16 and 417 were obtained by treatment of **11** and **12** with bromotrimethylsilane followed by alkaline or acidic hydrolysis, respectively.

Scheme 2

i: ((EtO)₂OP)₂CHLi, THF, -78°C (n=1, 55%; n=2, 80%). ii: 1) TBAF; 2) MsCl (n=1) or TsCl (n=2), NEt₃ iii: 6-ClGua, NaH (3 steps: n=1, 25%; n=2, 52%). iv: 1) TMSBr, CH₂CN; 2) NaOH 1N (n=1, 2steps: 42%) or 0.5N HCl (n=2, 2steps: 25%).

Synthesis of the unsaturated aldehydes 5 and 6 is depicted in scheme 3 starting respectively from commercially available 2-methylene-1,3-dichloropropane and 2-bromo-ethanol. On one hand, the dichloride was transformed into the alcohol derivative 13 in three steps by known methods. Then, the remaining hydroxy function of 13 was oxidized using N-methylmorpholine N-oxyde and a catalytic amount of tetrapropylammonium per-ruthenate (NMNO/TPAP)¹⁸ to afford the aldehyde 5. On the other hand, 2-bromo-ethanol was first protected as a TBDMS-ether and then alkylated with the sodium salt of diethylmalonate to give the malonate 14. The

reduction of the ester functions of 14 gave the dihydroxy derivative which, by a controlled monoacetylation reaction, afforded the monohydroxy derivative 15. The remaining hydroxy function of 15 was then oxidized using the NMNO/TPAP method. Interestingly, during the oxidation, the elimination of acetic acid occurred, affording the α,β -unsaturated aldehyde 6.

Scheme 3

i: 1) Et₄NOAc; 2) MeOH, K₂CO₃; 3) TBDMSCI, NEt₃ (3 steps: 35%). ii:1.5eq. NMNO, 0.05eq.TPAP, CH₂CI₂ (83%).

Br OH
$$\stackrel{i}{\longrightarrow}$$
 $\stackrel{EtO_2C}{\longleftarrow}$ OTBDMS $\stackrel{iii}{\longrightarrow}$ OTBDMS $\stackrel{iii}{\longrightarrow}$ OTBDMS $\stackrel{iii}{\longrightarrow}$ OTBDMS $\stackrel{iii}{\longrightarrow}$ OTBDMS

i: 1) TBDMSCI, NEt₃; 2) (EtO₂C)₂CH₂, NaH (2 steps: 44%). ii: 1) LAH, ether (40%); 2) Ac₂O, NEt₃, CH₂Cl₂ (61%). iii:1.5eq. NMNO, 0.05eq. TPAP, CH₂Cl₂ (77%).

The dienyl phosphonates 3 and 4 were evaluated as potential substrates of guanylate kinase¹⁹. Compound 4 proved to be an excellent substrate of the enzyme, being phosphorylated as efficiently¹⁹ as the natural substrate GMP (Table 1). The Vmax value of 4 is 3.3-fold lower than that of GMP. However, since the Km of 4 is also 3.4 lower than that of GMP, the efficiency¹⁹ of phosphorylation calculated for 4 is 102%. To our knowledge, compound 4 is the most efficient non-natural substrate ever reported for guanylate kinase. In contrast, its shorter chain analog 3 is a relatively poor substrate of guanylate kinase, being a 57-fold less efficient substrate than 4.

 Table 1.
 Comparison of kinetic parameters of GMP and dienyl phosphonates 3 and 4 for guanylate kinase

17 ± 2	100	100
57 ± 5	6.1	1.8
5 ± 2	30	102
	57 ± 5	57 ± 5 6.1

^a The kinetic parameters of 3 and 4 were determined at enzyme concentrations 20 and 5 times higher than that used for GMP, respectively.

^b For GMP, **3** and **4**, Km and Vmax values are means of 5, 2 and 3 separate determinations, respectively.

^c Vmax of GMP = 13 μmol . min⁻¹ . (mg protein)⁻¹. Standard deviations on mean Vmax values of **3** and **4** were 2 and 13%, respectively. Vmax are expressed in % of Vmax of GMP.

The anti-HIV-1 activity of dienyl phosphonates 3 and 4 was evaluated in the human T-lymphoid cell lines MT-4 and C-8166²⁰ infected with the HIV-1-RF strain (Table 2). The nucleoside analogues 3'-azido-3'-deoxythymidine (AZT), 2',3'-dideoxyinosine (ddI) and the acyclonucleotide analogue 9-[2-phosphonomethoxy)ethyl]adenine (PMEA) which are selective inhibitors of HIV replication were used as references²¹. In MT-4 and C-8166 cells, compound 4 inhibited HIV-1 replication with IC50 values of 49 and 8 μ M, respectively, and was therefore about 2.5- and 5-fold less potent than PMEA (Table 2). However, compound 4 was more than 5 times less cytotoxic to MT-4 cells than PMEA, thus showing an interesting selectivity index. In MT-4 cells, compound 3 did not inhibit HIV-1 replication at 100 μ M.

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Compound	EC ₅₀	CC ₅₀ (μM) ^c	
Compound	MT-4 cells	C-8166 cells	MT-4 cells
AZT	0.04 (± 0.01)	0.04 (± 0.03)	113 (± 36)
ddl	13 (± 10)	0.9 (± 0.4)	> 1000 *
PMEA	20 (± 10)	1.5 (± 1.0)	187 (±135)
3	> 100 *	n.d.	> 1000 *
4	49 (± 13)	8 (±2)	> 1000 **

Except for data maked with * or ** (from one or two experiments, respectively), data are mean values of at least three independent experiments (± SD); n.d., not determined.

Compounds 3 and 4 were also tested for activity against herpes simplex virus of type 1 (HSV-1, HF strain) and human cytomegalovirus (HCMV, AD 169 strain) grown in Vero and MRC-5 cells, respectively, by measuring inhibition of the virus-induced cytopathic effect as previously described ¹¹. Neither compound 3 nor 4 inhibited HCMV replication up to 250 μ M. In contrast, compound 3 inhibited HSV-1 replication with an IC50 value of 119 \pm 14 μ M (n = 2), i.e. similar to that of PMEA (117 \pm 38 μ M (n = 2)). Acyclovir, used as reference, had an IC50 value of 0.5 \pm 0.3 μ M (n = 2). Compound 4 was inactive against HSV-1 at 125 μ M. Compounds 3 and 4 were not toxic to Vero cells at least up to 250 μ M. In contrast to our observation, Harnden *et al.* ¹⁰ found that 3 showed no significant anti-HSV-1 (SC 16 strain) activity at 300 μ M. This discrepancy may be due to differences in uptake and/or intracellular activation (phosphorylation) of 3 between Vero and MRC-5 cells, the host cell system used by these authors.

^a Concentration required to inhibit HIV-1-induced cythopathic effect by 50% in MT-4 cells.

^b Concentration required to inhibit p24 viral antigen production by 50% in C-8166 cells.

^c Concentration required to reduce the viability of MT-4 cells by 50%.

In conclusion, we have shown that acyclic dienyl phosphonate derivatives of guanine can act as substrates of guanylate kinase. Compound 4 was found to be an excellent substrate of guanylate kinase and a significant inhibitor of HIV replication. In contrast, the shorter chain analogue 3 was a modest substrate of guanylate kinase and showed some activity only against HSV-1. These examples illustrate the potential of the dienyl phosphonate moiety as a surrogate of the phosphate group. In addition, the antiviral activity of such compounds could be related, at least in part, to their ability to serve as substrates for guanylate kinase.

Acknowledgements: We wish to thank A. Eggenspiller, A. Eschbach, J. Dulworth, T. Brennan for technical assistance and C. Reeb for typing this manuscript.

References and notes

- 1. Duke, A.E.; Smee, D.F.; Chernow, M.; Boehme, R.; Matthews, T.R. Antiviral Res. 1986, 6, 299.
- De Clercq, E.; Sakuma, T.; Baba, M.; Pauwels, R.; Balzarini, J.; Rosenberg, I.; Holy, A. Antiviral Res. 1987, 8, 261.
- 3. Pauwels, R.; Balzarini, J.; Schols, D.; Baba, M.; Desmyter, J.; Rosenberg, I.; Holy, A.; De Clercq, E. Antimicrob. Agents Chemother. 1988, 32, 1025.
- Kim, C.U.; Luh, B.Y.; Misco, P.F.; Bronson, J.J.; Hitchcock, M.J.M.; Ghazzouli, I.; Martin, J.C. J. Med. Chem. 1990, 33, 1207.
- 5. Duckworth, D.M.; Harnden, M.R.; Perkins, R.M.; Planterose, D.N. Antiviral Chem. Chemother. 1991, 2, 229.
- Yu, K.L.; Bronson, J.J.; Yang, H.; Patick, A.; Alam, M.; Brankovan, V.; Datema, R.; Hitchcock, M.J.M.; Martin, J.C. J. Med. Chem. 1993, 36, 2726.
- 7. Votruba, I.; Bernaerts, R.; Sakuma, T.; De Clercq, E.; Merta, A.; Rosenberg, I.; Holy, A. Mol. Pharmacol. 1987, 32, 524.
- Balzarini, J.; Hao, Z.; Herdewijn, P.; Johns, D.G.; De Clercq, E. Proc. Natl. Acad. Sci. USA 1991, 88, 1499.
- 9. Nevts, J. and De Clercq, E. Biochem. Pharmacol. 1994, 47, 39.
- 10. Harnden, M.R.; Parkin, A.; Parratt, M.J.; Perkins, R.M. J. Med. Chem. 1993, 36, 1343.
- 11. Navé, J-F.; Taylor, D.; Tyms, A.S.; Kenny, M.; Eggenspiller, A.; Eschbach, A.; Dulworth, J.; Brennan, T.; Piriou, F.; Halazy, S. Antiviral Res. 1995, 27, 301.
- 12. Miller, W.H.; Miller, R.L. J. Biol. Chem. 1980, 255, 7204.
- Smee, D.F.: Boehme, R.; Chernow, M.; Binko, B.P.; Matthews, T.R. Biochem. Pharmacol. 1985, 34, 1049.
- 14 Miller, W.H.; Miller, R.L. Biochem, Pharmacol. 1982, 31, 3879.
- 15. Minami, T.; Motoyoshiya, J. Synthesis 1992, p 333.

- The ¹H NMR (360 MHz, D₂O) spectrum of compound 3 was in good agreement with that reported by Harnden et al. ¹⁰
- 17. Compound 4: Analysis of $C_{11}H_{14}N_{5}O_{4}P$, HCl: calc. C: 38.00, H: 4.35, N: 20.14; found: C: 38.10, H: 4.31, N: 20.12. ^{1}H NMR (360 MHz, $D_{2}O$): δ 7.70 (s, 1H, H-8); 6.68 (dd, J_{HH} = 16 Hz, J_{HP} = 20 Hz, 1H, CH-4'); 6.09 (dd, J_{HH} = 16 Hz, J_{HP} = 14 Hz, 1H, CH-5'); 5.10 (s, 1H, H_{A} of C=CH₂); 4.78 (s, 1H, H_{B} of C=CH₂); 4.25 (t, J_{A} = 4 Hz, 2H, CH₂-1'); 2.75 (t, J_{A} = 4 Hz, 2H, CH₂-2').
- 18. Griffith, W.P.; Ley, S.V.; Whitcombe, G.P.; White, A.D. J. Chem. Soc. Chem. Comm. 1987, 1625.
- 19. Guanylate kinase (hog brain) was assayed according to Navé et al. Arch. Biochem. Biophys. 1992, 295, 253. Efficiency is defined as [(Vmax/Km)_{compound}/(Vmax/Km)_{GMP}] x 100. In this assay, the phosphorylation of 1 μmol of GMP in the presence of ATP ultimately results in the oxidation of 2 μmol of NADH. For compounds 3 and 4, 1 μmol of NADH was oxidized per μmol of compound phosphorylated. These stoichiometries were used for the calculation of the rates of phosphorylation.
- Anti-HIV-1 activity in MT-4 and C-8166 cells was determined by the MTT reduction dye method and p24 viral core antigen assay, respectively. For details, see reference 11.
- 21. Despite the fact that PMEA shows much lower anti-HIV activity than AZT *in vitro*²², this compound has proved to be a much more potent antiretroviral agent than AZT in monkeys acutely infected by the simian immunodeficiency virus^{23,24}.
- 22. Balzarini, J.; Naesens, L.; Herdewijn, P.; Rosenberg, I.; Holy, A.; Pauwels, R.; Baba, M.; Johns, D.G.; De Clercq, E. Proc. Natl. Acad. Sci. USA 1989, 86, 332.
- 23. Tsai, C.-C.; Follis, K.E.; Sabo, A.; Grant, R.F.; Bartz, C.; Nolte, R.E.; Benveniste, R.E.; Bischofberger, N. J. Infect. Dis. 1994, 169, 260.
- 24. Tsai, C.-C.; Follis, K.E.; Grant, R.F.; Nolte, R.E.; Bartz, C.R.; Benveniste, R.E.; Sager, P.R. J. Acquir. Immune Defic. Syndr. 1993, 6, 1086.

(Received in Belgium 26 September 1995; accepted 7 December 1995)